

## Erratum

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### Advanced back-cross QTL analysis of tomato. II. Evaluation of near-isogenic lines carrying single-donor introgressions for desirable wild QTL-alleles derived from *Lycopersicon hirsutum* and *L. pimpinellifolium*

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Due to a technical error, the tables 1–6 were unfortunately not included in the above-mentioned article.

We apologize for this error and print the missing tables.

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**Abstract** Improved-processing tomato lines were produced by the molecular breeding strategy of advanced backcross QTL (AB-QTL) analysis. These near-

isogenic lines (NILs) contained unique introgressions of wild alleles originating from two donor wild species, *Lycopersicon hirsutum* (LA1777) and *L. pimpinellifolium* (LA1589). Wild alleles targeted for trait improvement were selected on the basis of previously published replicated QTL data obtained from advanced backcross populations for a battery of important agronomic traits. Twenty three NILs were developed for 15 genomic regions which were predicted to contain 25 quantitative trait factors for the improvement of seven agronomic traits: total yield, red yield, soluble solids, brix × red yield, viscosity, fruit color, and fruit firmness. An evaluation of the agronomic performance of the NILs in five locations worldwide revealed that 22 out of the 25 (88%) quantitative factors showed the phenotypic improvement predicted by QTL analysis of the BC<sub>3</sub> populations, as NILs in at least one location. Per-location gains over the elite control ranged from 9% to 59% for brix × red yield; 14% to 33% for fruit color; 17% to 34% for fruit firmness; 6% to 22% for soluble-solids content; 7% to 22% for viscosity; 15% to 48% for red yield, and 20% to 28% for total yield. The inheritance of QTLs, the implementation of the AB-QTL methodology for characterizing unadapted germplasm and the applicability of this method to other crops are discussed.

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**Key words** Molecular breeding · Germplasm · Quantitative traits

**Table 1** Targeted quantitative factors from *L. hirsutum* LA1777 (H) and *L. pimpinellifolium* LA1589 (PM) and corresponding NILs. Targeted QTL are in lower case italics and targeted quantitative factors identified at subthreshold significance are in capitals (trait affected and marker locus) (Bernacchi et al. 1997b; Tanksley et al. 1996). Chr = chromosome. NIL = near isogenic line. SSC = soluble solids; BOS = viscosity Bostwick; YDT = total yield; YDR = red yield; BYR = brix red yield; FC = fruit color and FIR = fruit firmness. See Fig. 1 for cM lengths of introgressed fragments

Targeted factor	Chr	Donor	NIL
SSC-TG260/CT267	1	H	TA523
	1	H	TA1257
<i>ssc3.2</i> /BOS-TG417	3	H	TA1276
<i>ydt4.1</i> /YDR-CD59/BYR-CD59	4	H	TA1133
<i>fc4.2</i> / <i>byr4.1</i> /BOS-CT50/SSC-TG163/YDR-TG163	4	H	TA517
	4	H	TA1284
BYR-TG574/FIR-TG574/YDR-TG574	4	H	TA1138
<i>fc5.1</i> /BOS-TG441	5	H	TA1116
	5	H	TA1292
	5	H	TA1291
	5	H	TA1293
<i>ssc5.1</i> /FC-TG60/CT138	5	H	TA1118
	5	H	TA1117
	5	H	TA1297
<i>fc11.2</i>	11	H	TA1347
<i>fir3.1</i>	3	PM	TA515
<i>byr3.1</i> / <i>ssc3.1</i>	3	PM	TA524
	3	PM	94T872-13
	3	PM	94T868-24
SSC-CT101	5	PM	TA514
<i>vis9.1</i>	9	PM	94T873-30
	9	PM	TA534
FC-TG393	11	PM	TA516

**Table 2** Number of QTL/factors for which the corresponding NILs showed the expected phenotypic effect in at least one location. SSC = soluble solids; BOS = viscosity Bostwick; YDT = total yield; YDR = red yield; BYR = brix red yield; FC = fruit color; FIR = fruit firmness

	SSC	BOS	YDT	YDR	BYR	FC	FIR
Total # QTL/factors tested as NILs	6	4	1	3	4	5	2
# showing expected effect at $P \leq 0.1$	4	4	1	2	4	5	2
# showing expected effect at $P \leq 0.05$	4	3	0	1	3	4	2

**Table 4** QTL/factors classified according to significance of BC<sub>3</sub> detection and significance of NIL best representing the predicted effects. P = significance level of single point analysis (QTL) and of linear contrast between NIL and control (NIL Δ%). Names of QTL/factors are described in Table 1

Significance of NIL Δ%				
Significance of QTL/factor detection in BC <sub>3</sub>	ns	$0.1 \leq P < 0.01$	$0.01 \leq P < 0.001$	$0.001 \leq P < 0.0001$
$0.1 \leq P < 0.01$	<i>ssc5.1</i>	SSC-TG260/CT267 <i>ydt4.1</i> YDR-CD59 BYR-CD59 BOS-CT50 FC-TG60/CT138 BYR-TG574	SSC-CT101	BOS-TG441
$0.01 \leq P < 0.001$	YDR-TG574	YDR-TG163 FIR-TG574 <i>fc5.1</i>	BOS-417 FC-TG393 <i>byr3.1</i>	SSC-TG163
$0.001 \leq P < 0.0001$	<i>ssc3.1</i>	<i>fc4.2</i> <i>vis9.1</i>	<i>ssc3.2</i> <i>byr4.1</i> <i>fir3.1</i> <i>fc11.2</i>	

**Table 3** QTL-allelic effect ( $\Delta\%$ ) predicted for the NILs based on BC<sub>3</sub> data compared with observed performance of the NILs. Grouped data are QTL/factors statistics from BC<sub>3</sub> mapping (first row, in italics) and corresponding NIL/s statistics (subsequent rows). If QTL/factor did not reach the threshold significance in BC<sub>3</sub> only the trait affected is described (Bernacchi et al. 1997b). Statistics for QTL are  $\Delta\%$  = percent phenotypic change associated with the *L. hirsutum* (H) or *L. pimpinellifolium* (PM) allele; P = significance of the association (Bernacchi et al. 1997b). Statistics for NIL evaluation are  $\Delta\%$  = percent phenotypic difference relative to *L. esculentum* control E6203; P = significance of contrast NIL-E6203. Markers, in italics, are RFLP loci associated with QTL/factor. Introgression indicates the segments of H DNA contained in the NIL. IS = Israel; SP = Spain; CAP = California, Woodland; CAH = California, Stockton and JP = Japan. ns = not significant ( $P > 0.1$ ). na = not available in a given location. Avg.  $\Delta\%$  = average phenotypic effect associated to the H or PM allele, considering only deviations at  $P \leq 0.1$  irrespective of the direction of the difference. none sig = none of the contrasts were significant at  $P \leq 0.1$ . The NILs selected for best performance in the case of QTL/factors represented by more than one NIL are indicated in bold

Trait, QTL (BC <sub>3</sub> ) NIL	Donor	Marker Introgression	IS		SP		CAP		CAH		JP		Avg. $\Delta\%$
			$\Delta\%$	P	$\Delta\%$	P	$\Delta\%$	P	$\Delta\%$	P	$\Delta\%$	P	
<i>Soluble solids</i>	H	<i>TG260/CT267</i>	6	0.03	5	0.07	7	0.07					6.0
<b>TA523</b>		TG260-CT190	9	0.01	-3	ns	-3	ns	6	0.06	7	0.08	7.5
TA1257		CT190	2	ns	-4	ns	4	ns	1	ns	4	ns	ns
<i>Soluble solids, ssc3.2</i>	H	<i>TG417/CT243</i>	11	0.0009	12	0.0006	5	ns					11.5
TA1276		TG417-CT243	5	ns	-1	ns	1	ns	8	0.015	5	ns	8.3
<i>Viscosity (Bostwick)</i>	H	<i>TG417</i>			-2	0.013							-14.6
TA1276		TG417-CT243			2	ns	-9	0.1	-11	0.013	-22	no rep	-9.9
<i>Total Yield, ytd4.1</i>	H	<i>CD59</i>	16	0.03	20	0.078	9	0.1					14.8
TA1133		CD59	-12	0.1	-10	ns	20	0.1			28	0.1	12.0
<i>Red yield</i>	H	<i>CD59</i>	15.4	0.09	26	0.05	2	ns					20.7
TA1133		CD59	-14.2	0.02	3.5	ns	20	0.1	19.1	0.04	48.2	0.03	18.2
<i>Brix red yield</i>	H	<i>CD59</i>	17	0.03	15	ns	-5	ns					17.2
TA1133		CD59	-20	0.005	-5	ns	13	ns	11	ns	42	0.04	10.7
<i>Red yield</i>	H	<i>TG163</i>	-1	ns	-12	ns	33	0.006					32.5
<b>TA517</b>		TG305-CT50-TG163	-14	0.03	15	0.1	5	ns	-12	ns	1	ns	0.5
TA1284	H	CT50-TG163	-1	ns	-19	0.08	-32	0.02	-2	ns			-25.2
<i>Soluble solids</i>	H	<i>TG163</i>	3	ns	10	0.004	5	ns					9.6
<b>TA517</b>		TG305-CT50-TG163	9	0.01	10	0.008	6.31	0.014	22	<0.0001	13.8	0.002	12.2
TA1284		CT50-TG163	13	0.001	9	0.01	0.14	ns	5.5	0.1			9.2
<i>Brix red yield, bry4.1</i>	H	<i>TG163</i>	3	ns	6	ns	41	< 0.0001					41.0
<b>TA517</b>		TG305-CT50-TG163	-7	ns	267	0.01	13	ns	8	ns	16	ns	26.5
TA1284		CT50-TG163	11	0.1	-11	ns	-32	0.02	4.	ns			-10.2
<i>Fruit color, fc4.2</i>	H	<i>TG163</i>	46	0.0003	14	0.06	-3	ns					29.7
<b>TA517</b>		TG305-CT50-TG163	7	ns	0	ns	23	0.05	-14	0.1	33	0.03	14.0
TA1284		CT50-TG163	-9	ns	0	ns	-2	ns	7	ns			ns
<i>Viscosity</i>	H	<i>CT50</i>			12	0.04							12.1
TA517		TG305-CT50-TG163			-4	ns	-3	ns	-15	0.001			-15.3
<b>TA1284</b>		CT50-TG163			-13	0.02	-13	0.02	-6	ns			-12.8
<i>Red yield</i>	H	<i>TG574</i>	14	0.08	3	ns	37	0.007					26.5
TA1138		TG264-TG574-CT50	-3	ns	3	ns	16	ns					ns
<i>Brix red yield</i>	H	<i>TG574</i>	12	0.08	9	ns	34	0.01					22.7
TA1138		TG264-TG574-CT50	-1	ns	7	ns	24	0.08	12	0.1	13	ns	18.2

Table 3 Continued

Trait, QTL (BC <sub>3</sub> ) NIL	Donor	Marker Introgression	IS		SP		CAP		CAH		JP		Avg. Δ%
			Δ%	P	Δ%	P	Δ%	P	Δ%	P	Δ%	P	
<i>Firmness</i>	H	<i>TG574</i>	−9	ns	17	0.01	13	ns					17.0
TA1138		TG264-TG574-CT50	17	0.06	4	ns	2	ns	34	0.04	25	0.1	25.3
<i>Fruit color, fc5.1</i>	H	<i>TG441</i>	42	0.01	15	0.06	2	ns					28.4
<b>TA1116</b>		TG441-CT167-CD64	7	ns	17	0.04			−1	ns	8	ns	16.7
TA1291		TG441			0	ns	−2	ns	−1	ns			ns
TA1292		TG441			6	ns							ns
TA1293		CD64	7	ns	17	0.04	18	0.1	−1	ns			17.4
<i>Viscosity</i>	H	<i>TG441</i>			−12	0.05							−11.8
<b>TA1116</b>		TG441-CT167-CD64			−3	ns			−23	< 0.0001			−22.5
TA1291		TG441			6	ns	5	ns	15	0.001			14.7
TA1292		TG441			−7	ns							ns
TA1293		CD64			−11	0.053	1	ns	5	ns			−10.6
<i>Soluble solids, ssc5.1</i>	H	<i>TG69</i>	12	0.01	8	0.08	13	0.02					11.0
TA1118		TG69-CT138	3	ns	3	ns	−2	ns	4	ns	−1	ns	ns
<b>TA1117</b>		TG69-CT138	1	ns	−4	ns	−5	0.03	−1	ns	−1	ns	−5.4
TA1297		CT138	2	ns	−8	ns	−5	0.03	3	ns	1	ns	−5.4
<i>Fruit color</i>	H	<i>TG60/CT138</i>	31	0.06	5	0.01	6	0.052					14.0
TA1118		TG69-CT138	−4	ns	0	ns	13	ns	11	ns	25	0.1	25.0
<b>TA1117</b>		TG69-CT138	7	ns	6	ns	18	0.1	7	ns	8	ns	18.0
TA1297		CT138	−9	ns	−11	0.1	3	ns	−10	ns	8	ns	−11.1
<i>Fruit color, fc11.2</i>	H	<i>TG393</i>	46	0.0002	−3	ns	2	ns					46.2
TA1347		TG393	14	0.02	3	ns	0	ns	−20	0.02	8	ns	−2.6
<i>Soluble solids ssc3.1/3.2</i>	PM	<i>TG129/CD51/TG388</i>	7	0.04	7	0.0007	7	< 0.0001					7.1
<b>TA524</b>		TG246-TG129-CD51-TG388	−2	ns	−4	ns	−8	0.002	−4	ns	−1	ns	−7.9
94T872-13		TG246-TG129-CD51	3	ns	−11	0.03	na	na	−3	ns	−5	ns	−10.8
94T868-24		TG388	2	ns	−6	0.1	−5	0.04	−3	ns	−1	ns	−5.4
<i>Brix red yield, byr3.1</i>	PM	<i>CD51-TG388</i>	5	ns	18	0.001							18.2
<b>TA524</b>		TG246-TG129-CD51-TG388	9	0.1	2	ns	−8	ns	9	ns	59	0.005	34.1
94T872-13		TG246-TG129-CD51	25	0.001	−15	ns			0	ns	−7	ns	24.7
94T868-24		TG388	7	ns	−3	ns	−19	0.1	6	ns	8	ns	−19.4
<i>Firmness, fir3.1</i>	PM	<i>TG525</i>	40	0.0001									40.0
TA515		TG114-TG525-CT141-TG66	22	0.01	4	ns	2	ns	34	ns	−17	ns	22.2
<i>Soluble solids</i>	PM		−1	ns	1	ns	5.2	0.04					5.2
TA514		CT101	2.1	ns	11	0.005			−7	0.04	5	ns	1.70
<i>Viscosity, vis9.1</i>	PM	<i>TG421</i>			−13	0.0003							−13
<b>TA534</b>		TG421			−8	0.1	−7	0.1	−3	ns	−36	no rep	−7.5
94T873-30		TG421-CT74-TG551			−3	ns	−5	ns	−5	ns	−28	no rep	ns
<i>Fruit color</i>	PM		11	ns	23	0.009	1	ns					2.3
TA516		TG393-I2-TG36-TG546	18	ns	25	0.002	17	0.1	−15	0.07	25	0.07	12.9

**Table 5** QTL/factors classified according to the number of locations of BC<sub>3</sub> detection and the number of locations where corresponding NIL showed the predicted effect (NIL Δ%) (P ≤ 0.1). Names of QTL/factors are described in Table 1

Number Loc. QTL/factor detection in BC <sub>3</sub>	Number Loc. significant NIL Δ% in predicted direction				
	0	1	2	3	5
1		<i>fir3.1</i> SSC-CT101 BOS-TG441 <i>fc11.2</i> BYR-CD59 <i>byr4.1</i>	<i>byr3.1</i> YDR-TG163 BOS-CT50 BOS-417 <i>vis9.1</i>	FIR-TG574 FC-TG393	SSC-TG163
2	YDR-TG574	<i>fc5.1</i> <i>ssc3.2</i>	BYR-TG574 <i>fc4.2</i>	YDR CD59	
3	<i>ssc3.1</i> <i>ssc5.1</i>	FC-TG60/CT138	<i>ydt4.1</i>	SSC-TG260/CT267	

**Table 6** Percent phenotypic differences between NILs and the *L. esculentum* control for traits other than those predicted by QTL analysis results. Δ% = percent phenotypic difference. Only the best performing NIL for each target are considered. IS = Israel; SP = Spain; CAP = California, Woodland; CAH = California, Stockton and JP = Japan. FC = fruit color; BYR = brix × red yield; YDR = red yield; FIR = fruit firmness; AFW = average fruit weight; SSC = soluble solids; BOS = viscosity Bostwick; YDT = total yield. Loc = location. The original association

between the introgressed markers and any given trait are given under BC<sub>3</sub> association (Bernacchi et al. 1997b) as follows: if a QTL was detected with significance in the BC<sub>3</sub>, its name is given as well as the estimated phenotypic effect associated with the wild allele; if an association failing to reach QTL significance was detected, the location in which it was detected is given in brackets, preceded by a negative sign (–) if the H or PM allele was associated with inferior performance or a positive sign (+) if H or PM allele improved the trait

NIL	Trait	NIL deviations from E6203 at P < 0.1								BC <sub>3</sub> mapping
		Loc	Δ%	Loc	Δ%	Loc	Δ%	Loc	Δ%	
TA253	FC	IS	+15							<i>fc1.3</i> – 33%, (–CAP) (–CAP) (–IS, –CAP) (+CAP)
	BYR	CAH	+22	JP	+48					
	YDR	JP	+38							
	FIR	CAH	+46							
	AFW	CAH	–10	CAP	–20	SP	–22			
TA1276	FC	CAP	–4	JP	–10					(–SP, +CAP) (–SP) (–IS, –SP, –CAP) (–IS, –SP)
	FIR	CAH	–23	SP	–12					
	YDR	IS	–11							
	AFW	CAH	–15	CAP	–12	IS	–15	SP	+11	
TA1133	BOS	CAH	–9	CAP	–8	SP	–20			(–CAP)
	SSC	CAH	–7	CAP	–6	IS	–6	SP	–7	
	AFW	CAH	–14	IS	–15	SP	–18			
TA517	pH	SP	+2	JP	+1.5					(–SP, –CAP) (–SP) (–SP)
	YDT	IS	–16							
	FIR	SP	–12	JP	–23					
	AFW	CAH	–25	CAP	–21	IS	–21	SP	–20	
TA1284	YDT	CAP	–31							(–SP)
	AFW	CAH	–15	IS	–15	SP	–16			
TA1138	BOS	CAH	+12	CAP	+13	JP	+21			(–SP, –CAP)
	SSC	CAH	+7	CAP	+6					
	AFW	CAH	–12	CAP	–12					
	FC	SP	–8							
TA1116	SSC	CAH	+9	IS	+8					+25
	FIR	CAH	+53	IS	+22	JP				
	AFW	CAH	–20	SP	–21					
TA1117	FIR	JP	–23	SP	–17					<i>fir5.1</i> – 32% (–SP) <sup>a</sup> (–IS, –SP)
	BYR	IS	–20	SP	–20					

**Table 6** Continued

NIL	Trait	NIL deviations from E6203 at $P < 0.1$								BC <sub>3</sub> mapping
		Loc	$\Delta\%$	Loc	$\Delta\%$	Loc	$\Delta\%$	Loc	$\Delta\%$	
TA1347	FIR	CAH	+27							
	BYR	JP	+53	SP	-24					
	YDR	JP	+51							
	BOS	CAH	-15							(-SP)
	SSC	CAP	-7	SP	-7	IS	+5			
	FC	CAP	-21							(+IS)
	AFW	CAH	-11							(-IS, -SP)
TA524	BOS	CAH	+10							(-SP)
	YDR	JP	+60	IS	+11					(+SP)
	FIR	JP	+33							
	AFW	CAH	-15	SP	-16					(-CAP, -IS, -SP)
TA516	BYR	SP	-24							(+IS)
	SSC	CAP	-5							(+CAP, +SP)
	AFW	CAH	-8	CAP	-25	IS	-16	SP	+12	(-CAP, -IS, -SP)
TA514	BYR	IS	+18							
	FC	IS	+36							(+SP)
	AFW	JP	+34							
	FIR	SP	-12	JP	-33					
TA534	YDR	JP	+30	SP	+29					(+IS) <sup>a</sup>
	BYR	SP	+18							
	FIR	CAH	+34	SP	+13					
	SSC	SP	-8							(-SP) <sup>a</sup>
	FC	CAH	-22	CAP	-16	SP	-11			(-SP)
TA515	FIR	CAH	+53	CAP	-23					(+IS)
	AFW	SP	+20	CAH	-13	CAP	-32			(-IS)

<sup>a</sup>A QTL or a subthreshold association was detected for the RFLP marker adjacent to introgressed segment